



## Marine ecosystem monitoring in the sea-ice region of Lützow-Holm Bay, off Syowa Station, during the icebreaker *Shirase* cruise of the 59th Japanese Antarctic Research Expedition

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**Abstract:** As part of the monitoring program of the Japanese Antarctic Research Expedition (JARE), marine ecosystem monitoring in the sea-ice region of Lützow-Holm Bay, off Syowa Station, has been routinely carried out since JARE-52 (2010/11 season) to investigate biological production and mechanisms in relation to sea-ice. Water temperature, salinity, chlorophyll-*a* concentration, macro-nutrients (nitrate, nitrite, phosphate, and silicate), and zooplankton data were collected at six stations in various sea-ice environments: fast-ice, pack-ice, and ice-free open ocean. The use of an “ice-fence” protected observation equipment (e.g., plankton nets and CTDs) from damage due to sea-ice. This report provides the data obtained during the icebreaker *Shirase* cruise of the 59th Japanese Antarctic Research Expedition in the 2017/18 season, and is the second report for the marine ecosystem monitoring in the sea-ice region from the first report of the 2016/17 season in this journal.

## 1. Background and Summary

The Southern Ocean marine ecosystems have been changing dramatically due to the effects of global climate change. Among the most obvious physical changes are the regionally contrasting variations in the extent and seasonality of sea-ice<sup>1</sup>. The Antarctic sea-ice zone is an ecologically important region of the Southern Ocean. The seasonal fluctuation of sea-ice formation and melting events has a significant effect on the life cycle of many organisms living in this region. However, since it is difficult to access by existence of sea-ice in these areas, the accumulation of biological knowledge is extremely poor compared to the open ocean region until now.

The powerful ice-breaking capability of the current Japanese icebreaker *Shirase* (Japan Maritime Self-Defense Force) raises the possibility of marine observations in the fast-ice and/or permanent ice zone. The ship-based marine ecosystem monitoring program for the sea-ice region of Lützow-Holm Bay, off Syowa Station, East Antarctica, began with the 52nd Japanese Antarctic Research Expedition (JARE-52) during 2010/11 season<sup>2</sup>. The aim of this program is to investigate biological production and mechanisms in relation to sea-ice.

This report presents the water temperature, salinity, macro-nutrients, phytoplankton chlorophyll-*a* concentrations, and zooplankton monitoring data obtained during a cruise by the icebreaker *Shirase* as part of the JARE-59 (February 2018). Similar datasets collected previously by JARE have been published in JARE Data Reports<sup>2, 3, 4, 5, 6, 7, 8, 9, 10, 11</sup>, and this journal<sup>12</sup>. This report is the second report on the marine ecosystem monitoring in the sea-ice region in this journal.

## 2. Sampling Location

Seawater and plankton samples were collected at six monitoring stations (A, B, C, D, E, and BP) in Lützow-Holm Bay in various sea-ice environments: fast-ice, pack-ice, and ice-free open ocean ([Fig. 1](#) and [Table 1](#)). At the time of sampling, Station A was in the multi-year fast-ice zone, Station B was in the first-year fast-ice zone, Stations C and D were in the pack-ice zone, and Stations E and BP were in the open-ocean zone.

## 3. Methods

### 3.1. Protection of the observation equipment

Oceanographic observations in sea-ice areas have the problem that the existence of sea-ice makes it difficult to use conventional equipment such as plankton net. We devised the “ice-fence” which guards observation equipment from sea-ice ([Fig. 2](#))<sup>13</sup>. Lützow-Holm Bay is a region where fast-ice cover persists during summer and heavy sea-ice states continue. Therefore, we modified the ice-fence several times in order to carry out observation under heavy sea-ice conditions, and succeeded in protecting equipment from the damage by sea-ice<sup>14</sup>.

### 3.2. Temperature and salinity

Vertical profiles of temperature and salinity were determined at six stations using a conductivity-temperature-depth (CTD) memory probe (SBE19 plus, Sea-Bird Scientific) attached to a water sampler with six 4-L bottles (SBE 55 ECO, Sea-Bird Scientific). The data were downloaded from the CTD to a laptop computer immediately after each cast. The CTD sensor was calibrated by Sea-Bird Scientific during the last boreal summer. The salinity data in this report were not corrected by the bottle salinity data by the salinometer.

### 3.3. Chlorophyll *a*

Vertical water samples were collected over the upper 100 m of the water column using six Niskin bottles attached to the water sampler, and surface water was collected using a plastic bucket. Samples for chlorophyll *a* measurement were collected in two dark bottles (ca. 300 mL) for bulk measurements using a glass-fiber filter (Whatman, GF/F) and size-fractionated measurements to determine the phytoplankton size composition using a Whatman nuclepore membrane filter (pore size: 10 and 2  $\mu\text{m}$ ) and a Whatman GF/F. The filters were immediately soaked in N, N-dimethylformamide<sup>15</sup>, and the pigments were extracted for more than 24 hours. The samples were stored ( $-18^{\circ}\text{C}$ ) until onboard analysis. Concentrations of chlorophyll-*a* were determined fluorometrically<sup>16</sup> with an onboard fluorometer (10-AU; Turner Design, Sunnyvale, CA, USA). The fluorometer was calibrated against a chlorophyll-*a* standard (Fujifilm Wako Pure Chemical Corp., Osaka, Japan) at a laboratory on-land before the cruise, using a spectrophotometer and the specific absorption coefficient of chlorophyll-*a*<sup>17</sup>. For a fluorometer calibration result, see Makabe *et al.* (2020)<sup>18</sup>.

### 3.4. Macro-nutrients

Seawater to determine macro-nutrient contents was collected at the time when seawater was sampled for chlorophyll-*a*. The sampled water was stored ( $-18^{\circ}\text{C}$ ) until analysis in a land laboratory. Nutrient concentrations were measured with a QuAAtro 2-HR system (provided by BL Tec K.K.). The nutrient (nitrate, nitrite, silicate, and phosphate) concentration measurements followed the methods of Hydes *et al.* (2010)<sup>19</sup>. A detailed description of the processing technique is presented in Shimada *et al.* (2021)<sup>20</sup>. The coefficients of variation calculated from five replicates of nitrate, nitrite, silicate, and phosphate were 0.15%, 0.23%, 0.18%, and 0.18%, respectively.

### 3.5. Zooplankton sampling

Zooplankton samples were collected using a closing net (mouth diameter 0.75 m, mesh size 100  $\mu\text{m}$ ) at stations A to D, and a twin NORPAC net (mouth diameter 0.45 m, mesh size 315  $\mu\text{m}$  and 100  $\mu\text{m}$ ) at stations E and BP. To prevent sea-ice from entering the net, an “ice-fence” is employed and

the net was closed as it reaches the surface<sup>13</sup>. The net was equipped with a flow-meter to estimate the volume of water filtered, and was hauled vertically from a depth of 150 m to the surface at stations where the bottom is deeper than 150 m, or from 5 m above the bottom to the surface at stations where the bottom is shallower than 150 m. All samples are fixed immediately in seawater with 5% buffered formalin.

## 4. Data Records

### 4.1. Temperature, salinity, macro-nutrients, and chlorophyll-*a* concentration

All measurements are presented in two data files, named “JARE59\_Sea-ice\_CTD”, and “JARE59\_Ch1&Nuts”. The fields in the datasheets in the two files are:

**CTDPRS** – the pressure (dbar) of the data collected

**CTDTMP** – the temperature (degree C)

**CTDCND** – the conductivity (s/m)

**CTDSAL** – the salinity

**THETA** – the potential temperature (degree C)

**SIGT** – the sigma-t ( $\text{kg m}^{-3}$ )

**CRUISE** – the cruise code of the vessel

**SHIP** – the name of the ship on which the sampling was conducted

**STNNBR** – the name of the station on which the sampling was conducted

**DATE** – the sampling date (DD/MM/YYYY)

**TIME** – the sampling time (UTC)

**LATITUDE** – the decimal latitude of the sampling station (negative value for South)

**LONGITUDE** – the decimal longitude of the sampling station (positive value for East)

**DEPTH** – the sampling depth (m)

**SIG0** – the sigma- $\theta$  ( $\text{kg m}^{-3}$ )

**CHL BULK** – total chlorophyll-*a* concentration ( $\mu\text{g l}^{-1}$ )

**CHL 10UM** – composition of chlorophyll-*a* in  $>10 \mu\text{m}$  fraction (%)

**CHL 2UM** – composition of chlorophyll-*a* in 2 – 10  $\mu\text{m}$  fraction (%)

**CHL GF/F** – composition of chlorophyll-*a* in  $<2 \mu\text{m}$  fraction (%)

**NITRAT** – concentration of nitrate ( $\mu\text{mol l}^{-1}$  or  $\mu\text{mol kg}^{-1}$ )

**NITRIT** – concentration of nitrite ( $\mu\text{mol l}^{-1}$  or  $\mu\text{mol kg}^{-1}$ )

**SILCAT** – concentration of silicic acid ( $\mu\text{mol l}^{-1}$  or  $\mu\text{mol kg}^{-1}$ )

**PHSPHT** – concentration of phosphate ( $\mu\text{mol l}^{-1}$  or  $\mu\text{mol kg}^{-1}$ )

## 4.2. Zooplankton dataset

Zooplankton monitoring datasets are presented at data file named “JARE59\_Sea-ice\_Zooplankton” in three data sheets, species/taxa list, abundance data and wet weight data. The fields in the dataset are:

**JARE number** – the JARE number of this sampling season

**Ship name** – the name of the ship on which the sampling was conducted

**Station number** – the name of the station on which the sampling was conducted

**Latitude** – the decimal latitude of the sampling station (negative value for South)

**Longitude** – the decimal longitude of the sampling station (positive value for East)

**Sampling season** – two-year Antarctic season based around the austral summer, e.g. “2017-2018” runs from December 2017 to March 2018

**Sampling year** – the sampling date year

**Sampling month** – the sampling date month

**Sampling day** – the sampling date day

**Sampling time** – the sampling date time (UTC)

**Sampling depth** – the depth of sampling tow

**Mesh size** – the mesh size of plankton net

**Estimated volume of water filtered** – the estimated volume of water filtered using a flow-meter

**Abundance** – the abundance of each species/taxa

**Total abundance** – total abundance of all zooplankton in a sample

**Number of species/taxa** – the number of species/taxa in a sample

**Wet weight** – the wet weight of each category

**Total wet weight** - total wet weight of all zooplankton in a sample

## 5. Technical Validation

### 5.1. Zooplankton identification

Zooplankton were identified to the lowest practical taxonomic level, generally to species or genus, using a stereo-microscope. Copepodite stages of copepod species, calytopis stages and furcilia stages of euphausiid species were subdivided from the adults. The nauplius stages of *Rhincalanus gigas* (Copepoda: Calanoida) were distinguished from other calanoid nauplii by their large size and morphology. Zooplankton abundance was converted to individuals per cubic meter.

The species list for this dataset was checked using the Taxon Match of the World Register of Marine Species (WoRMS: <http://www.marinespecies.org/index.php>) name validation tool. WoRMS is an open-access inventory of all marine species, being >90% complete<sup>21</sup>. The tool performs a cross-check of the spelling and taxonomic status of species against the WoRMS database; it returns

standard taxonomic information with valid names.

## 5.2. Wet weight measurement

Processing of samples was carried out according to the four-step procedure outlined below and shown in [Fig. 3](#).

Step 1: the large-sized zooplankton (more than 10 mm in size) were sorted for the whole sample in the laboratory. Zooplankton were classified into seven categories, and counting the number of individuals and the wet weight each category was measured using an electronic balance (Sartorius Quintix124-1SJP, readability 0.1 mg).

Step 2: all other species (<10 mm in size) were counted from 1/2 to 1/32 aliquots of the whole sample, and identified to the lowest taxonomic level, generally species or genus, level, using a stereo-microscope. We used a Motoda box-type plankton sample splitter to subdivide a sample. While sorting and counting this size fraction, the wet weight of zooplankton typically reached more than 10 mg per aliquot.

Step 3: given that the wet weight of zooplankton of 1 mm size or less were hard to sort, this fraction was estimated by using conversion factors listed in [Table 2](#).

Step 4: the weights obtained in steps 1-3 were summed to give a total wet weight. Zooplankton abundance was converted to  $\text{mg m}^{-3}$ . For a detailed description of zooplankton processing for wet-weight measurements, see Ukai *et al.* (2014)<sup>30</sup>.

## 6. Figures

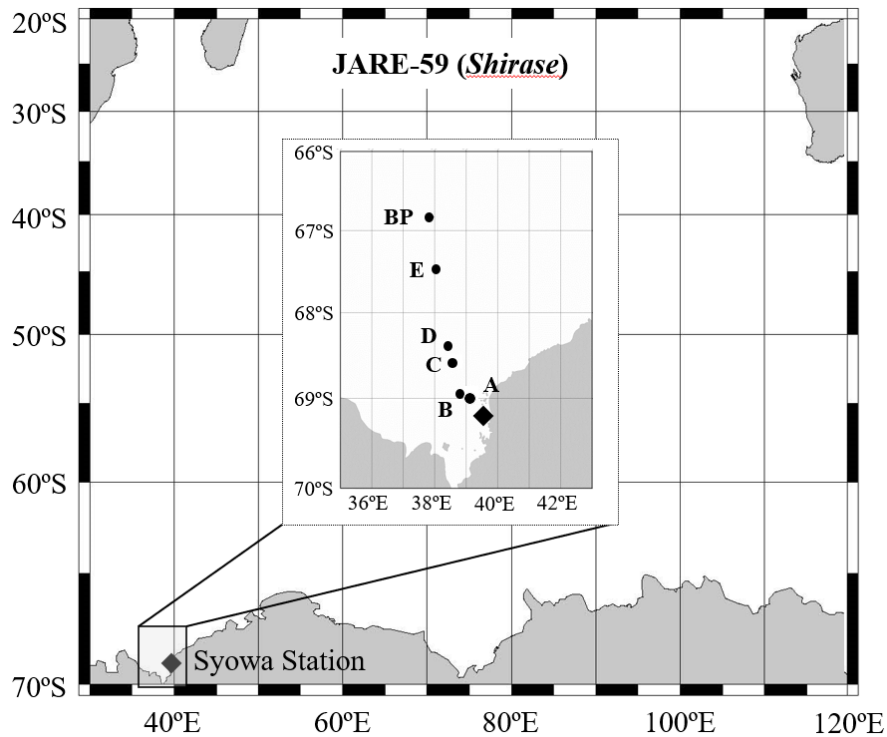


Fig. 1. Location of the sampling sites at Lützow-Holm Bay in February 2018.

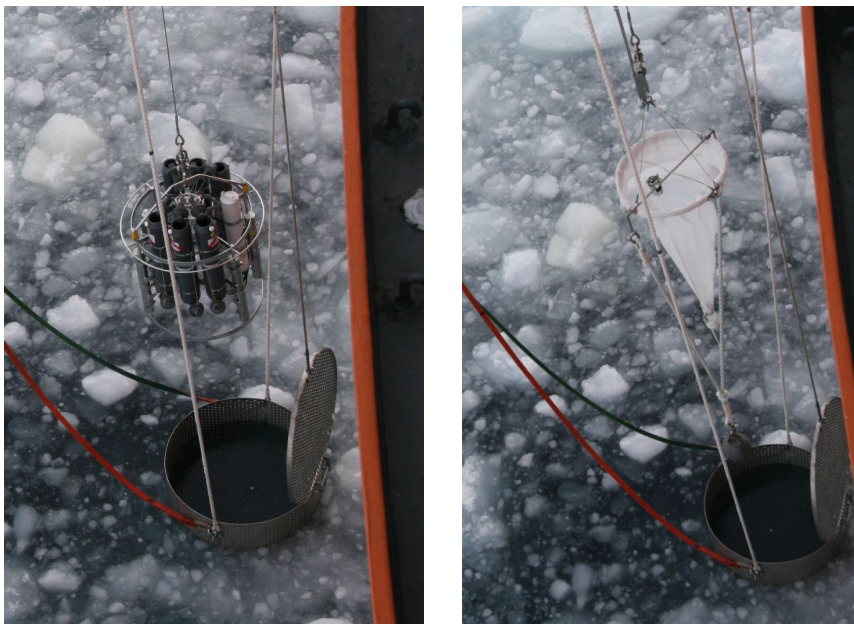


Fig. 2. CTD with a water sampler (left) and plankton net (right) used in the monitoring program in the sea-ice region. We used the “ice-fence” which guards observation equipment from sea-ice. Modified Figure 7 of Takahashi et al. (2012)<sup>13</sup>.

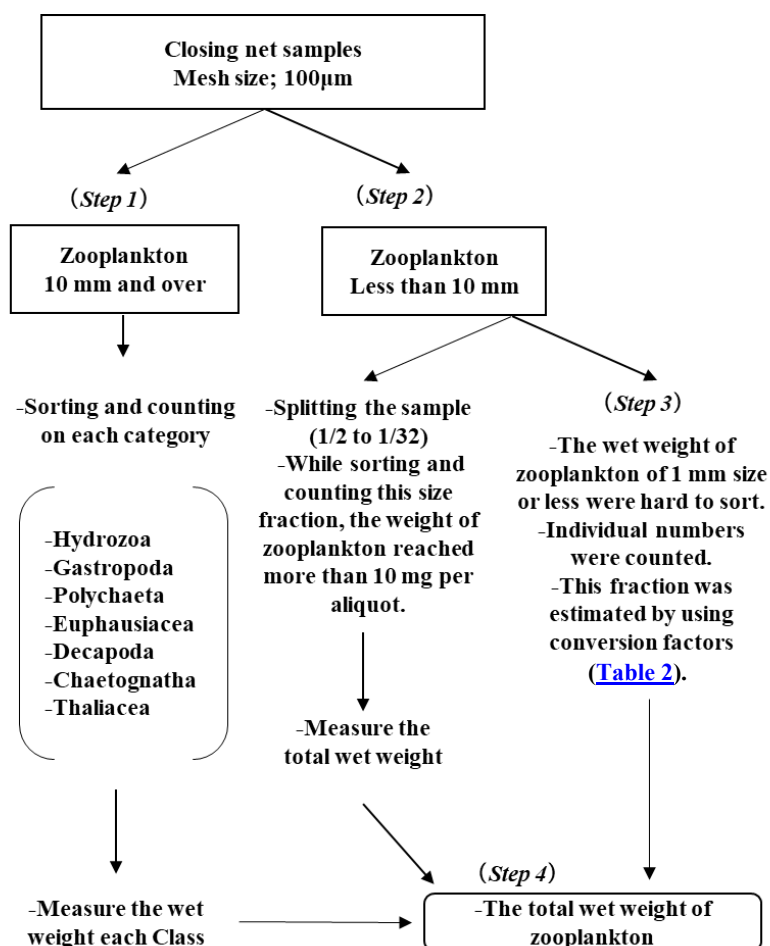


Fig. 3. Four-step procedure used to measure and to estimate the wet weight of NORPAC net samples.

## 7. Tables

Table 1. Sampling information of CTD cast and closing & NORPAC net at six stations

Station	Date (UTC)	CTD cast			Closing & NORPAC net sampling		
		Time (UTC)	Latitude (°S)	Longitude (°E)	Time (UTC)	Latitude (°S)	Longitude (°E)
A	11/02/2018	09:51	-69.00001	39.16700	10:14	-69.00004	39.16659
B	11/02/2018	12:07	-68.83130	38.75266	12:23	-68.83146	38.75235
C	15/02/2018	09:55	-68.62573	38.58092	10:17	-68.62577	38.58092
D	15/02/2018	12:50	-68.42052	38.45751	13:14	-68.42051	38.45757
E	16/02/2018	07:35	-67.49832	38.01831	08:11	-67.50333	38.00657
BP	16/02/2018	11:02	-66.83147	37.84770	11:28	-66.83189	37.82929



### Author contributions

K.T. Takahashi performed the processing of zooplankton samples and writing of the manuscript. T. Odate directed the JARE-59 monitoring program. R. Makabe and S. Takao carried out the nutrient analysis in the land laboratory and analyzed the sensor data.

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